



Original article

Walatimine (Vanillyl Butyl Imine): A New Ketimine from Schiff Base Synthesis and Evaluation of its Antioxidant, Antibacterial and Antifungal Properties

Olawale Hakeem Oladimeji*, Joy Adesoji Olukoju, Samuel Ogayi Ogbu, Uforo Joseph Ubobo and Emmanuel Edet Attih

Department of Pharmaceutical and Medicinal Chemistry, Faculty of Pharmacy, University of Uyo, Uyo, Nigeria

ARTICLE INFO

Received 26 April 2023
Revised 24 May 2023
Available Online 30 June 2023

ACADEMIC EDITOR
Dr. Taleuzzaman

*CORRESPONDING AUTHOR
Olawale Hakeem Oladimeji,
Department of Pharmaceutical
and Medicinal Chemistry, Faculty of
Pharmacy, University of Uyo, Uyo,
Nigeria

ABSTRACT

Background of study: Vanillin is a white crystalline compound which is a phenolic aldehyde with a balsamic flavour. It is widely obtained from vanilla bean-pods amongst very many other sources. This compound and some of its derivatives possess diverse activities including anticancer, anti-inflammatory, antioxidant, antibacterial and antifungal amongst others. Hence, the import of this study.

Objectives: The growing concerns about the deleterious actions of free radical oxygenated species (FROS) in the human body have become a huge concern to the scientific world. These chemical species continually devastate the human cells, tissues and organs leading to different pathophysiological conditions and neurodegenerative disorders. Also, the noticeable microbial resistance to antibiotics and antifungal drugs have prompted the search for lead compound(s) with the aim of chemically modifying its/their molecular structure(s) or synthesizing other compounds from reactions involving them such as Schiff base synthesis. The search for novel pharmaceutically active compounds with the aim of ameliorating these conditions led to the choice of vanillin.

Methodology: Vanillin was subjected to the Schiff base synthesis. It was reacted separately with amines (aromatic and aliphatic amine) leading to the bases in the presence of acid. The melting points, refractive indices and optical rotations of the vanillin and the Schiff bases were obtained. The antioxidant activity (IC₅₀) of the lead compound and bases was determined employing the DPPH (2, 2-diphenyl-1-picrylhydrazyl hydrate) bench-top assay test. Comparison of the obtained antioxidant activities was done to determine if any improvements could be noticed in the synthesized bases. Also, the agar-in-hole diffusion method was adopted for screening vanillin and the synthesized bases against *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Candida albicans* for antibacterial and antifungal activities respectively.

Results: The identities of the synthesized Schiff bases have been revealed to be vanillyl butyl imine (W) a new ketimine referred to by the trivial name-Walatimine and vanillin-2,4-dinitrophenyl hydrazone (Vdnp) respectively

using a combination of physico-chemical determinations and IR spectral technique. Vanillin and (W) demonstrated marginal antioxidant activity of IC_{50} of 0.52 and 0.50 $\mu\text{g/mL}$ respectively while Vdnp gave a remarkably significant IC_{50} of 0.48 and which compare favourably with 0.46 $\mu\text{g/mL}$ elicited by Vitamin C (a standard antioxidant drug). The antibacterial and antifungal activities elicited by both W and Vdnp were concentration dependent. Furthermore, Vdnp was comparably more bacteriostatic than W against the test bacteria though it was inactive against *Ps. aeruginosa*. However, vanillyl butyl imine (W) was slightly more anti-candidal against *C. albicans* than vanillin-2,4-dinitrophenyl hydrazone (Vdnp).

Conclusion: The results from this study indicate that vanillin-2,4-dinitrophenyl hydrazone (Vdnp) obtained from the condensation reaction of vanillin and an aromatic amine affords a comparably better antioxidant activity than W obtained from vanillin reacting with an aliphatic amine. Both W and Vdnp demonstrated remarkable antibacterial and antifungal activities hence, these two synthesized Schiff bases can be lead candidate compounds in the search for newer and more efficacious antioxidant and antimicrobial agents and in further structural activity relationship studies (SARS) and as well as in formulation studies in drug development.

Keywords: Vanillin; Vanillyl butyl imine; Vanillin-2,4-dinitrophenyl hydrazone; Antioxidant; Antibacterial; Antifungal

Abbreviations

W: Vanillyl butyl imine (Walatimine); Vdnp: Vanillin-2, 4-dinitrophenyl hydrazone; DPPH: 2, 2-Diphenyl-1-picrylhydrazyl hydrate; RSA: Radical Scavenging Activity; PI: Percentage Inhibition; IC_{50} : Concentration at which 50 % of DPPH is scavenged or inhibited; NCTC: National Collection of Type Cultures, Central Public Health Laboratory, Colindale Avenue, London NW9, UK; ATCC: American Type Culture Collection, Washington, DC; NCYC: National Collection of Yeast Cultures, UK.

Introduction

Vanillin is one of the most popular and widely used aromatic flavouring compounds found as a glycoside in the fruits of natural vanilla. The main source of vanilla is the bean or pod of the tropical vanilla orchid, *Vanilla planifolia* but also found in *Vanilla fragrans*, *Vanilla pompona* and *Vanilla tahitensis*. It was discovered amongst the Aztecs of Mexico by the Spaniards who introduced it to Europe in 1520 and is now cultivated around the world in countries such as Mauritius, Seychelles, Madagascar, Java, Ceylon, Tahiti, Guadeloupe, Martinique and Indonesia amongst many others. In 1874, Ferdinand Thompson and Wilhelm Haarman synthesized vanillin from coniferin, a glycoside of isoeugenol found in pine bark. Currently, synthetic vanillin is used as an intermediate in the chemical and pharmaceutical industries for the

manufacture of herbicides, antifoam agents, drugs such as papaverine, L-methyldopa, L-dopa, trimethoprim and also in the production of fragrances and flavoring agents [1-4]. Many derivatives of vanillin have shown good antibacterial properties against gram (+) and gram (-) microbes. Vanillin is also known to have antioxidant properties due to their strong free radical scavenging properties and derivatives obtained from its reduction, O-demethylation, oxidation and acetylation respectively gave between marginal and moderate antioxidant activities using the bench-top assay with DDPH reagent [5]. The increasing application of antibiotics in healthcare has thrown up the risk of growing resistance as multi-drug resistance bacterial pathogens are rising and there is a growing need for stronger and more effective antibiotics. Hence, the need to explore for novel alternatives. Similarly, for fungal infections therapy which also suffers from resistance occasioned by multi-drug resistant fungi thereby making a search for newer antifungal drugs/agents inevitable. Schiff base was first reported by Hugo Schiff in 1864. It is also known as imine which is a pharmacophore containing the azomethine group ($-\text{HC}=\text{N}-$). It can also be referred to ketimine or aldimine or hydrazone or aziridine. The ketimines in particular are used as anesthetic agents in animals after their introduction in 1970 by Federal Drug Administration (FDA) in America. This drug is employed by emergency responders on agitated patients or to calm them and as well as in treating depression and suicidal

thoughts in patients. Imines or Schiff bases are the products formed in condensation reactions of ketones or aldehydes with amines and generally take place in the presence of acid or base or heat. The formation of a Schiff base is a reversible reaction that is usually completed by the separation of product or removal of water or both. It is best carried out at mild acidic pH. It can be hydrolyzed back to their aldehydes or ketones and amines by aqueous acid or base. The mechanism involves the addition of a nucleophile (amine) to the carbonyl group. The amine reacts with the aldehyde or ketone to form an unstable compound called carbinolamine which undergoes dehydration catalyzed by acid, thus loses water. Schiff bases are one of the most widely used organic compounds in the production of pigments, dyes, catalysts and polymer stabilizers. Reports exist which show that these compounds exhibit a wide range of biological activities including antioxidant, antibacterial, antifungal, anti-malarial, antidepressant, anti-proliferative, anti-inflammatory, anticancer, antipyretic, anti-diabetic and antidepressant properties [6-10]. Schiff bases with anthracene and pyrene units have reportedly been found to be antibacterial against *Bacillus cereus*, *Escherichia coli* and *Pseudomonas aeruginosa* in *in-vitro* studies while benzothiazole-based Schiff ligands were bacteriostatic against *Staphylococcus aureus* [11]. Furthermore, cinnamyl Schiff bases have shown antifungal activities against *Candida albicans* and *Aspergillus fonsecaea* [12] while phenylenediamine containing Schiff bases and those with metallic ions such as zinc, chromium, copper and manganese have demonstrated radical scavenging activity [13]. Consequently, this present research was designed to synthesize Schiff bases using vanillin, a compound with an aldehydic group (-HC=O) and two different amines separately. One, a straight chain aliphatic amine (butyl amine) while the other was aromatic (2,4-dinitrophenyl hydrazine). The synthesized Schiff bases were screened for antioxidant activity (IC₅₀) using the DPPH reagent and the agar-in-hole diffusion method was employed in determining the anti-bacterial and antifungal sensitivity properties. Comparison of results obtained was done with values given by vanillin and the synthesized Schiff bases and as well as the positive controls such as Vitamin C (antioxidant drug), chloramphenicol (antibiotic) and fluconazole (antifungal drug) with a view to determining if any improvements could be noticed in the targeted biological activities of the synthesized Schiff bases.

Materials and Methods

Reagents/chemicals

Both DPPH (2, 2-diphenyl-1-picryl hydrazyl hydrate) and vanillin were sourced from Tianjin Kernel Chemical Reagent Company, China and Sigma Aldrich Chemicals, Germany respectively while chloramphenicol, fluconazole and Vitamin C tablets were obtained from Fidson Healthcare PLC, Nigeria. Reagents namely, acetic acid (glacial), acetone, butyl amine, ethanol, ethyl acetate, 2,4-dinitrophenyl hydrazine, hydrochloric acid, methanol, n-butanol, n-hexane, petroleum-ether and toluene were purchased as AnaLAR Grade Chemicals from British Drug House Chemicals Limited, Poole, England.

Solubility/Dissolution tests for vanillin

Vanillin (0.03 g) was added to 3 mL of each of the following solvents namely, dilute HCl, distilled water, ethyl acetate, ethanol, petroleum ether, n-hexane, n-butanol and methanol separately and observation was made for complete dissolution (solubility) or otherwise.

Determination of melting point

Vanillin (0.04 g) was filled to a quarter of the length of a micro-capillary tube and the melting point determined [14] using an Electro-thermal Melting Point apparatus (Electro-thermal Engineering Limited, England).

Synthesis of vanillyl butyl imine (Walatimine)

This Schiff base was synthesized as described by [15] with slight modifications. Butyl amine (3 mL) was added to 30 mL of glacial acetic acid to give a mixture. To this mixture was a solution of vanillin (3 g in 10 mL glacial acetic acid) added drop wise whilst stirring and the reaction mixture (green colour) was heated under reflux (glass chamber) for 8 h. At the completion of reaction, the yellow product mixture obtained was allowed to cool over crushed ice possibly for crystallization to occur. The mixture in the absence of the formation of crystals was then concentrated down *in vacuo* on a rotary evaporator (920H-Flaswk, Switzerland) at 40°C to dryness (Figure 1,2).

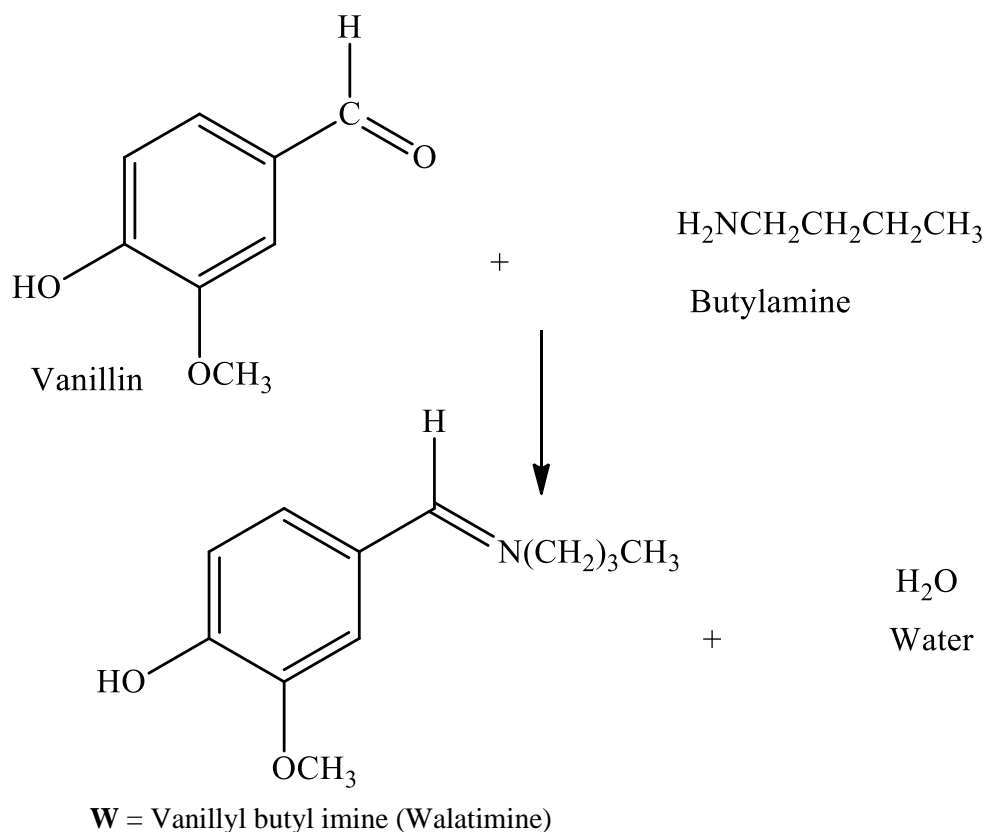


Figure 1: Synthesis of Walatimine.

Determination of optical rotation and refractive indices of vanillin and the synthesized vanillin Schiff bases

This is done by using a polarimeter (ADP-220, Bellingham Stanley, England) and a refractometer (WAY-15, Abbe, England). Each sample (0.05 g) was dissolved in methanol (10 mL). The tube of the polarimeter was filled with distilled water and the machine subsequently zeroed. The tube was then refilled with 5 mL of sample and the optical rotation and was measured at the wavelength (λ) of sodium D line (589.3nm) at 20.5°C. Similarly, the refractive index of sample was obtained on a refractometer at the wavelength (λ) of sodium D line (589.3 nm) at 20.5°C [16,17].

Antioxidant activity

Spectrophotometric determination of antioxidant activity using DPPH reagent Substances which are

capable of donating electrons or hydrogen atoms can convert the purple-coloured DPPH radical (2, 2-diphenyl-1-picrylhydrazyl hydrate) to its yellow-coloured non-radical form; 1, 1-diphenyl-2-picryl hydrazine [18,19]. This reaction can be monitored by spectrophotometry.

Preparation of calibration curve for DPPH reagent

This experiment was carried out as described by both [5,20] with some modifications. DPPH (4 mg) was weighed and dissolved in methanol (100 mL) to produce the stock solution (0.004% w/v). Serial dilutions of the stock solution were then carried out to obtain the following concentrations viz, 0.0004, 0.0008, 0.0012, 0.0016, 0.0020, 0.0024, 0.0028, 0.0032 and 0.0036% w/v. The absorbance of each of the sample was taken at λ_m 517 nm using the Ultra-Violet Spectrophotometer (Jenway 6405, USA). This machine was zeroed after an absorbance had been taken with a solution of methanol without DPPH which served as the blank (Figure 3).

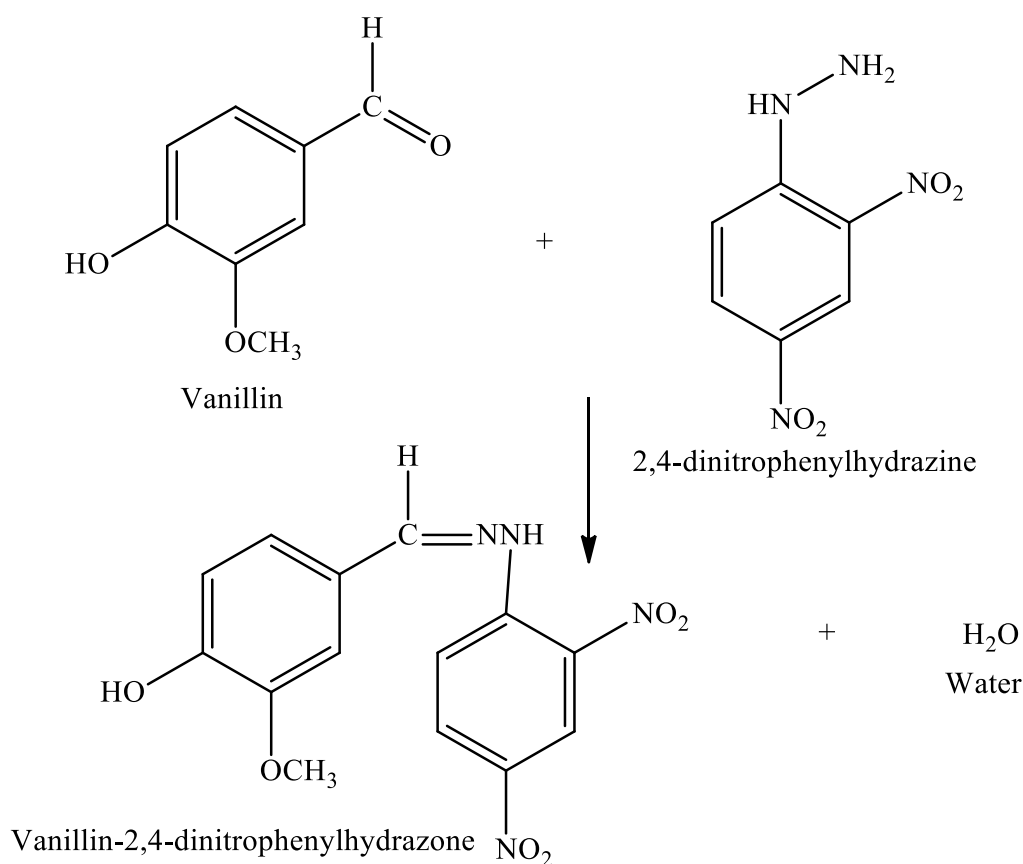


Figure 2: Synthesis of vanillin -2,4-dinitrophenyl hydrazine (Vdnp).

Determination of the antioxidant activity of vanillin, synthesized Schiff bases and Vitamin C

2 mg of sample was mixed with 50 mL of methanol. Serial dilutions were carried out to obtain the following concentrations; 0.0004 mg mL⁻¹, 0.0008 mg mL⁻¹, 0.0012 mg mL⁻¹, 0.0016 mg mL⁻¹ and 0.0020 mg mL⁻¹ using methanol. 5 mL of each concentration was incubated with 5 mL of 0.004 % w/v methanolic DPPH solution for optimal analytical accuracy. After an incubation period of 30 minutes in the dark at room temperature (25 ± 2°C), observation was made for a change in the colour of the mixture from purple to yellow. The absorbance of each of the samples was then taken at λ_m 517 nm. The Radical Scavenging Activity (RSA %) or Percentage Inhibition (PI %) of free radical DPPH was thus calculated:

$$RSA \% (PI \%) = [(A_{\text{blank}} - A_{\text{sample}}) / A_{\text{blank}}] \times 100$$

A_{blank} is the absorbance of the control reaction (DPPH solution without the test sample) and A_{sample} is the absorbance of DPPH incubated with the sample. Vanillin /synthesized Schiff base / Vitamin C concentration providing 50 % inhibition (IC₅₀) was calculated from a graph of inhibition percentage against the concentration of the vanillin/ synthesized Schiff base /Vitamin C [21-23]. Vitamin C was used as a standard antioxidant drug.

Antimicrobial Tests

The micro-organisms used in this study, namely; *Bacillus subtilis* (NCTC 8432), *Staphylococcus aureus* (NCTC 4532), *Escherichia coli* (NCTC 1065), *Pseudomonas aeruginosa* (ATCC 4675) and *Candida albicans* (NCYC 2436) were clinically isolated from specimens of diarrheal stool, abscesses, necrotizing fasciitis, osteomyelitis, urine, wounds and vaginal swabs obtained from the Medical Laboratory, University of Uyo Health Centre, Uyo. The clinical isolates were

collected in sterile bottles, identified and typed by convectional biochemical tests [24-25] and then refrigerated at -5°C at the Microbiology and Parasitology Unit, Faculty of Pharmacy prior to use. The hole-in-plate agar diffusion method was used observing standard procedures for bacterium and fungus respectively. The inoculum of each micro-organism was introduced into each petri-dish (Pyrex, England). Cylindrical plugs were removed from the agar plates by means of a sterile cork borer (Pyrex, England) to produce wells with diameter of approximately 5.00 mm. The wells were equidistant from each other and the edge of the plate [26][27]. Concentrations of 20 mg mL^{-1} of vanillin, 10 mg mL^{-1} and 20 mg mL^{-1} of synthesized Schiff bases were introduced into the wells. Also, different concentrations of $5\text{ }\mu\text{g mL}^{-1}$ chloramphenicol (Fidson Healthcare Chemicals, Nigeria), 1 mg mL^{-1} of fluconazole (Fidson Healthcare Chemicals, Nigeria) and aqueous methanol (1:1) were introduced into separate wells as positive and negative controls respectively [28-31]. The experiments were carried out in triplicates. The plates were left at room temperature for 2 h to allow for diffusion. The plates were then incubated at $37\pm 2^{\circ}\text{C}$ for 24 h. Zones of inhibition were measured in millimetre (mm).

Thin-layer chromatography of vanillin and synthesized Schiff bases

A portion of each solid sample (0.03 g) dissolved in methanol (2 mL) was applied on a 20 cm x 10 cm silica gel analytical plate (Merck, Germany) and then developed in a toluene : acetone : water (10:20:1) mixture in a chromatographic tank until optimal separation was observed [5]. The retardation factor (R_F) was then computed thus:

$$R_F = \frac{\text{distance moved by spot}}{\text{distance moved by solvent front}}$$

Infra-red spectroscopy of vanillin and synthesized Schiff bases

Each sample (0.03 g) was analyzed for IR characteristics using the FTIR 84005 Spectrophotometer (Shimadzu, Japan).

Ultra-violet/visible spectroscopy of vanillin and synthesized Schiff bases

A portion of each sample (0.03 g) was analyzed for UV/VS absorption characteristics using the Jenway 6405 UV/VS Spectrophotometer (Table 1).

Table 1: Preparation of calibration curve for DPPH reagent at λ_{max} 517 nm.

Concentration	Average absorbance (± 0.004)
0.0004	0.069
0.0008	0.169
0.0012	0.237
0.0016	0.321
0.002	0.391
0.0024	0.445
0.0028	0.537
0.0032	0.652
0.0036	0.703
Blank Absorbance of 0.004% w/v DPPH reagent: (0.911)	

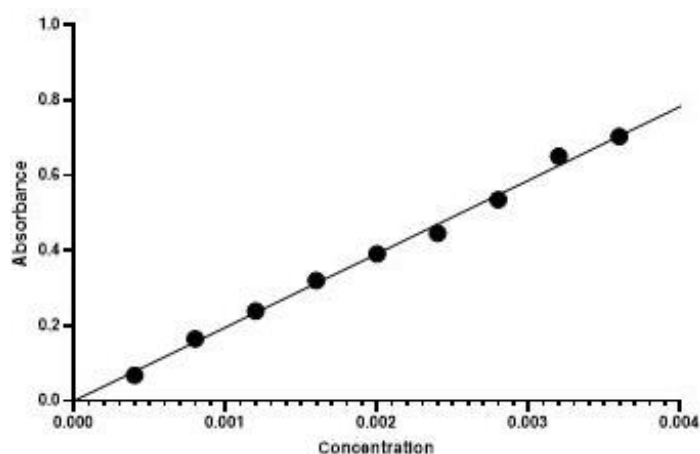


Figure 3: The calibration curve of DPPH reagent.

Table 2: Absorbance of samples incubated with DPPH at different concentrations at λ max 517 nm (Blank absorbance of 0.004% DPPH reagent: 0.911) (\pm 0.004).

Sample	0.0008 mg mL ⁻¹	0.0016 mg mL ⁻¹	0.0024 mg mL ⁻¹
Vitamin C	0.066	0.064	0.062
Vanillin	0.272	0.27	0.269
W	0.251	0.23	0.164
Vdnp	0.082	0.073	0.071

Table 3: Radical scavenging activity (percentage inhibition %) of samples at different concentrations and IC₅₀ of samples (\pm 0.02).

Sample	0.0008 mg mL ⁻¹	0.0016 mg mL ⁻¹	0.0024 mg mL ⁻¹	IC ₅₀ (μ g mL ⁻¹)
Vitamin C	92.65	92.76	92.86	0.46
Vanillin	70.14	70.36	70.47	0.52
W	72.23	74.25	82.11	0.5
Vdnp	91.2	91.78	91.99	0.48

Vanillin: C₈ H₈ O₃; mol. wt. (152.15 g/mol); white crystals (solid); m.pt. (81-83 °C); [n]_D²⁰ (1.5776); [α]_D²⁰ (0 °); λ _{max} (246 nm); R_F (0.64); FTIR (cm⁻¹): 1145 (-C-O-C), 1587 (-Ar-C=C), 1664 (-C=O), 2917 (-CH stretching) and 3289 (-Ar-OH).

Vanillyl butyl imine (Walatimine) W: C₁₂ H₁₇ NO₂; mol. wt. (207.08 g/mol); yellow compound; [n]_D²⁰ (1.5880); [α]_D²⁰ (0 °), λ _{max} (412 nm); R_F (0.81); FTIR (cm⁻¹): 875 and 921 (finger print region, alkyl bending

mode), 1145 (-C-O-C), 1583 (Ar-C=C), 1691 (-HC=N, of imine), 2917 (-CH stretching) and 3288 (-Ar-OH).

Vanillin-2,4-dinitrophenyl hydrazone (Vdnp): C₁₄ H₁₂ N₄ O₆; mol. wt. (332.27 g/mol); pale brown resin; [n]_D²⁰ (1.5984); [α]_D²⁰ (0 °); λ _{max} (308 nm); R_F (0.60); FTIR (cm⁻¹): 1145 (-C-O-C), 1355 (-O₂N group), 1567 and 1601 (-Ar-C=C), 2917 and 2918 (-CH stretching) and 3291 (-Ar-OH).

Table 4: Antibacterial sensitivity screening of vanillin and synthesized Schiff bases at different concentrations on test microbes in MeOH/ H₂O (1:1) (\pm 0.01 mm)

Test microbe	Vanillin 20 mg L ⁻¹	W 10 mg mL ⁻¹	W 20 mg L ⁻¹	Vdnp 10 mg mL ⁻¹	Vdnp 20 mg mL ⁻¹	Chloramphenicol 5 μ g mL ⁻¹	MeOH/ H ₂ O (1:1)
<i>B. subtilis</i> (NCTC 8432)	32.21	16.23	30.16	21.26	38.31	46.01	5.00
<i>S. aureus</i> NCTC 4532)	29.23	15.11	32.19	25.32	35.22	45.23	5.00
<i>E. coli</i> (NCTC 1065)	30.65	18.20	29.12	26.22	37.21	47.45	5.00
<i>Ps. aeruginosa</i> (ATCC 4675)	27.24	19.45	30.64	5.21	5.10	26.12	5.00

Note: The zone diameter recorded is zone of inhibition + size of cup (zone of inhibition +5.00) mm.

Table 5: Antifungal sensitivity testing of vanillin and synthesized Schiff bases at different concentrations on test microbes in MeOH/H₂O(1:1) (\pm 0.01 mm).

Test microbe	Vanillin 20 mg L ⁻¹	W 10 mgmL ⁻¹	W 20 mg L ⁻¹	Vdnp 10 mgmL ⁻¹	Vdnp 20 mgmL ⁻¹	Fluconazole 1 mgmL ⁻¹	MeOH/H ₂ O (1:1)
<i>C. albicans</i> (NCYC 2436)	22.21	15.32	28.23	14.12	24.34	43.54	5.00

Note: The zone diameter recorded is zone of inhibition + size of cup (zone of inhibition +5.00) mm.

Discussion

Spectroscopic analyses

Vanillin is a white crystalline compound with a pleasant and balsamic fragrance. Some monographic determinations were done in this study where the identity, purity, integrity and suitability substance were established. The compound was observed to be soluble in ethanol, ethyl acetate, n-butanol, n-hexane, methanol and water when heated. However, it was insoluble in petroleum ether, dilute hydrochloric acid and dilute sulphuric acid. The observations and the determined melting point and refractive index values are consistent with those in literature and those obtained in an earlier study [5]. The UV absorption characteristic at λ_{\max} (246 nm) indicates the presence of electron clouds over -Ar-C=C, -OH, -OCH₃ and -HC=O pharmacophores while retardation factor R_F (0.64) shows that vanillin is moderately polar and likewise retarded on the silica gel. The IR spectrum of vanillin shows absorptions at 1145, 1587, 1664, 2917 and 3289 cm⁻¹ which are diagnostically characteristic of -C-O-C, -Ar-C=C, -C=O, -CH and -Ar-OH groups respectively (Table 1-5). The condensation reaction between vanillin and butyl amine led to the synthesis of a yellow compound which has been identified by as vanillyl butyl imine by a combination of physico-chemical determinations and the IR spectral technique and hereby coded as W. It has a slightly pleasant flavour. Comprehensive data-based library searches of organic compounds were done and hence it is safe to infer that the compound is novel and hereby referred to by a trivial name Walatimine. It is a Schiff base which belongs to the class of compounds known as ketimines or aldimines. These compounds are obtained by condensation reactions between amines and carbonyl moieties such as ketones or aldehydes in the presence of an acid or base and under heat. The UV absorption at λ_{\max} (412 nm) which is comparably higher than that of the vanillin shows the presence of electrons de-localized over -Ar-C=C, -OCH₃, -HC=N (imine) and -Ar-OH

chemical species while the retardation factor R_F (0.81) shows that this Schiff base is comparably non-polar on account of the butyl group (-CH₂)₃CH₃ which makes the compound more lyphophilic and hence weakly retarded on an apparently hydrophobic silica gel plate. The IR spectral matrix of W shows stretchings at 875 and 921, 1145, 1583, 1691, 2917 and 3288 cm⁻¹ which are characteristically diagnostic of alkyl bending modes (finger print region), -C-O-C (ether linkage), -Ar-C=C, -HC=N (imine), -CH and -Ar-OH respectively. Also, in a somewhat similar fashion, the condensation reaction between vanillin and 2,4-dinitro phenyl hydrazine resulted in a pale brown product which has been identified to be vanillin-2,4-dinitro phenyl hydrazone (Vdnp) using similar techniques as mentioned above. This Schiff base belongs to the class of compounds referred to as hydrazones or aziridines. The UV absorption at λ_{\max} (308 nm) is higher than that of vanillin at 246 nm but less than that of W at 412 nm. However, the electron densities are found over -O₂N, -Ar-C=C, -OCH₃, -HC=N and -Ar-OH chemical species inherent in the compound while the retardation factor R_F (0.60) is indicative of this compound being more adsorbed onto the silica gel than the vanillin at 0.64. This observation is not surprising because the two nitro groups (on the second benzene ring) and -OH (on the first benzene ring) both contribute to making the synthesized hydrazone more polar hence, more greatly retarded on the silica gel analytical plate. Furthermore, the IR spectral matrix of vanillin-2,4-dinitro phenyl hydrazone (Vdnp) shows absorptions at 1145, 1355, 1587, 1601, 2917, 2918 and 3291 cm⁻¹ which correspond to the presence of -C-O-C (ether linkage), -O₂N (nitro group), -Ar-C=C, -CH bending modes and -Ar-OH chemical entities respectively. Moreover, the presence of the two O₂N (nitro group) being electron withdrawing chemical entities will predispose the characteristic -Ar-C=C and -CH absorptions to higher IR peaks (especially in the second benzene ring) as can be seen in the IR spectrum of the synthesized hydrazone. The determinations of physical parameters are important in identifying compounds. Physical

constants such as refractive index and optical rotation are used in the qualitative and quantitative analyses of substances. Also, these parameters are employed to confirm the purity, identity, integrity of active substances and as well as monitor the progress of reactions. In this study, both physical parameters were measured at the wavelength (λ) of Na-D light (589.3 nm) and a temperature of 20.5 °C. In addition, the refractive index of a substance is an indication of the number, type of atoms and chemical groups (species) in the substance. Each atom or group in the substance contributes to its refractivity which adds eventually to the refractive index of the substance. Furthermore, refractive index can be used to monitor the progress of chromatographic separation by measuring the refractive indices of the effluent solvents employed. The pro-drug, vanillin and both synthesized Schiff bases gave refractive indices of 1.5776, 1.5880 and 1.5984 respectively. Furthermore, it was observed that all the compounds demonstrated optical rotation $[\alpha]_D^{20}$ of 0° implying that the compounds do not have chiral centers hence, are optically inactive. In addition, none of these compounds will demonstrate laevorotation (-) (ability of a compound to rotate plane of light in anticlockwise direction) or dextro-rotation (+) (ability of a compound to rotate plane of light in clockwise direction) [16][17].

Antioxidant assays

In absorption spectrophotometry, it is germane that a calibration curve be prepared for the reagent to be employed in the assays. Hence, this was done for DPPH (2, 2-diphenyl-1-picryl hydrazyl hydrate) reagent with the aim of ascertaining its purity and suitability for use in the antioxidant determinations. The Beer-Lambert's Law is for such determinations [19,32]. The calibration curve obtained indicates that the underlying principles behind the afore mentioned Law were fulfilled as the curve (Figure) shows a straight line which passes through the origin. The reduction of the DPPH radical was determined by taking its absorption at a wavelength of λ_m 517 nm. It was observed that the absorbance of DPPH decreased as the concentration of added free radical scavenger (vanillin /Schiff base/Vitamin C) increased which suggested that the DPPH reagent was being reduced (Table 2), On the other hand table 3 shows radical scavenging activity (RSA %) or percentage inhibition (PI %) and the computed IC_{50} values of vanillin /Schiff base / Vitamin C. The RSA % is an indicator or a measure of the antioxidant activity of vanillin / Schiff base / Vitamin C. Interestingly, vanillin, and vanillyl butyl imine (W) both demonstrated moderate antioxidant activity (IC_{50}) of

0.52 and 0.50 $\mu\text{g mL}^{-1}$ respectively. However, W being more lyphophilic because of the presence of the butyl group $-(\text{CH}_2)_3\text{CH}_3$ has a slightly better activity than the vanillin because the 4-carbon aliphatic long chain will ensure that the compound gets to the active or allosteric sites relatively faster where the pharmacological action of anti-oxidation is effectuated. Vanillin-2,4-dinitro phenyl hydrazone (Vdnp) showed a significantly remarkable antioxidant activity at 0.48 $\mu\text{g mL}^{-1}$ which compares favourably with that of Vitamin C (a standard antioxidant drug) at 0.46 $\mu\text{g mL}^{-1}$ It could be inferred that the product (Vdnp) of condensation reaction between vanillin and an aromatic amine such as 2,4-dintro phenyl hydrazine slightly enhances the antioxidant activity of vanillin than that it forms with a straight chain aliphatic amine such as butyl amine. Vanillin like most low molecular weight phenolic compounds possess weak or in some instances moderate antioxidant properties [5]. This sweet-smelling fragrant compound also has been proven to demonstrate anticancer, anti-inflammatory and neurone-protective activities [33]. Hydrazones have been reported to possess anti-infective activity [34]. Aside from the DPPH test for determining the antioxidant activity of compounds include the hydrogen peroxide, nitric oxide, conjugated diene, superoxide, phosphomolybdenum, peroxy nitrile and xanthine oxidase assay methods amongst many others [35,36].

Antibacterial sensitivity tests

The microbes employed in the sensitivity tests reflected the antibacterial spectrum encompassing two (2) gram positive bacteria namely, *B. subtilis* and *S. aureus* and two (2) gram negative bacterial species namely, *E. coli* and *Ps. aeruginosa*. The results displayed on the Table 4 show that vanillin was remarkably suppressive or bacteriostatic of all the four microbes at 20 mg m L⁻¹ while the antibacterial activities recorded by both W (vanillyl butyl imine) and Vdnp (vanillin-2,4-dinitro phenyl hydrazone) were concentration-dependent at 10 and 20 mg m L⁻¹ respectively as reflected on Table 4. The higher the concentration, the better the activity. Generally, both bases demonstrated good antibacterial activities as can be seen on Table 4. However, W was noticeably and remarkably suppressive of *Ps. aeruginosa* at both concentrations while Vdnp was inactive. This observation was surprising because *Ps. aeruginosa*, a gram negative bacterium is well known for its unique resistance to antibacterial agents. This resistance is believed to be due to the nature of the cell envelope of the organism which unlike gram positive organisms possess a sophisticated three-layered

envelope which does not allow the permeation of external agents [37]. It can be inferred from these observations that both bases could be promising lead compounds in the search for newer antibacterial agents for treatment and management of bacterial infections.

Antifungal activity

The antifungal screening was done with *C. albicans* and generally the three compounds displayed somewhat remarkable anti-candidal activities at the concentrations tested. Vanillin was tested at 20 mg mL⁻¹ while W and Vdnp were screened at both 10 and 20 mg mL⁻¹ respectively. Similarly, the antifungal activity demonstrated by the two Schiff bases are concentration-dependent as can be seen on Table 5. However, it is noted that W was slightly more anti-candidal than Vdnp.

Conclusion

This study reports for the first time the synthesis of vanillyl butyl imine (W), a new ketimine from Schiff base synthesis. It has been given the trivial name Walatimine. This compound demonstrated a moderate antioxidant activity (IC₅₀) of 0.50 as vanillin at 0.52 µg mL⁻¹. However, vanillin-2,4-dinitro phenyl hydrazone (Vdnp) gave a significant antioxidant activity (IC₅₀) of 0.48 µg mL⁻¹ which compare favourably with that shown by Vitamin C at 0.46 µg mL⁻¹. Vanillyl butyl imine (W) demonstrated remarkable antibacterial activities against the bacteria tested including *Ps. aeruginosa*, an organism noted for its resistance to anti-infective agents. Also, it was slightly more anti-candidal than vanillin-2,4-dinitro phenyl hydrazone (Vdnp). It is noteworthy that both W and Vdnp could be lead compounds in the search for newer antioxidant and antimicrobial agents and in further structural activity relationship studies (SARS) and as well as in formulation studies in drug development.

Acknowledgements

The authors acknowledge the assistance of the Department of Pharmaceutical and Medicinal Chemistry, University of Uyo, Nigeria for the use of its Jenway 6405UV/VS. The efforts of Mr E. U. Akpan of the Department of Pharmaceutical Microbiology and Biotechnology, University of Uyo, Nigeria is hugely appreciated. We are also grateful to university of Ibadan, Nigeria for the use its IR-FTIR 8400 Spectrophotometer (Shimadzu, Japan).

Compliance with Ethical Standards

Not applicable.

Funding

The authors did not receive any financial sponsorship for the project.

Conflict of Interest

The author declares no conflict of interest.

References

1. Burri J, Graf M, Lambelet P, Löliger J. Vanillin: more than a flavouring agent-a potent antioxidant. *Journal of the Science of Food and Agriculture*. 1989;48(1):49-56.
2. Hocking MB. Vanillin: synthetic flavoring from spent sulfite liquor. *Journal of chemical education*. 1997 Sep 1;74(9):1055.
3. Davidson PM, Naidu AS. Phytophenols. In: Naidu, A.S., *Natural Food Antimicrobial Systems*. CRC Press LLC, Boca Raton, Washington, DC, 2000, pp. 265-294.
4. Walton NJ, Mayer MJ, Narbad A. Vanillin. *Phytochemistry*. 2003 Jul 1;63(5):505-15.
5. Oladimeji HO, Idiong ES, Joseph UA, Anah VO, et al. A cycloalkanol from derivatization studies on vanillin; Evaluation of the antioxidant activity of the obtained derivatives. *Amer. J. Quantum Chem. & Molecular Spectroscopy*, 2022, 6(1), 1-7
6. Dhar DN, Taploo CL. Schiff bases and their applications. *J. Sci. & Ind. Res.*, 1982, 41(8), 501-506
7. Przybylski P, Huczynski AW, Pyta KK, Brzezinski B, Bartl F. Biological properties of Schiff bases and azo derivatives of phenols. *Current Organic Chemistry*. 2009;13.
8. Yadamani S, Neamati A, Homayouni-Tabrizi M, Beyramabadi SA, Yadamani S, Gharib A, Morsali A, Khashi M. Treatment of breast cancer by using low frequency electromagnetic fields and Mn (II) complex of a Schiff base derived from the pyridoxal. *The Breast*. 2018 Oct 1;41:107-12.
9. Ansari MJ, Aldawsari MF, Zafar A, Soltani A, Yasir M, Jahangir MA, Taleuzzaman M, Erfani-Moghadam V, Daneshmandi L, Mahmoodi NO, Yahyazadeh A. In vitro release and cytotoxicity study of encapsulated sulfasalazine within LTSP micellar/liposomal and TSP micellar/niosomal nano-formulations. *Alexandria Engineering Journal*. 2022 Dec 1;61(12):9749-56.

10. Mohanty D, Gilani SJ, Zafar A, Imam SS, Kumar LA, Ahmed MM, Jahangir MA, Bakshi V, Ahmad W, Eltayib EM. Formulation and Optimization of Alogliptin-Loaded Polymeric Nanoparticles: In Vitro to In Vivo Assessment. *Molecules*. 2022 Jul 13;27(14):4470.
11. Mishra N, Kumar K, Pandey H, Anand SR, Yadav R, Srivastava SP, Pandey R. Synthesis, characterization, optical and anti-bacterial properties of benzothiazole Schiff bases and their lanthanide (III) complexes. *Journal of Saudi Chemical Society*. 2020 Dec 1;24(12):925-33.
12. Magalhães TF, da Silva CM, Dos Santos LB, Santos DA, Silva LM, Fuchs BB, Mylonakis E, Martins CV, de Resende-Stoianoff MA, de Fátima Â. Cinnamyl Schiff bases: Synthesis, cytotoxic effects and antifungal activity of clinical interest. *Letters in Applied Microbiology*. 2020 Nov 1;71(5):490-7.
13. Wu W, Xuan Y, Yin JL, Shujie L. Synthesis and Characterization of a Novel o-Vanillin Schiff Base Derivatives and Their Complexes with Zn (II). *Synthesis and Reactivity in Inorganic, Metal-Organic, and Nano-Metal Chemistry*. 2014 Jul 3;44(6):877-80.
14. Jahangir MA, Khan S, Singh AD, Muheem A, Soni A, Taleuzzaman M. Nanophytomedicine in clinical management: An introductory evidence-based review. *Journal of Pharmaceutical Research Science & Technology [ISSN: 2583-3332]*. 2022 Feb 7;6(1):26-37.
15. Azam F, Singh S, Khokhra SL, Prakash O. Synthesis of Schiff bases of naphtha [1, 2-d] thiazol-2-amine and metal complexes of 2-(2'-hydroxy) benzylideneaminonaphthothiazole as potential antimicrobial agents. *Journal of Zhejiang University Science B*. 2007 May;8:446-52.
16. Oladimeji OH, Njinga S, Abdullahi ST. Evaluation of antioxidant activity of obtained derivatives of vanillin. *Journal of Pharmaceutical Research Science & Technology [ISSN: 2583-3332]*. 2022 Feb 7;6(1):17-25.
17. Olaniyi AA. Principles of drug quality assurance and pharmaceutical analysis. Ibadan: Mosuro Publishers. 2000.
18. Huang G, Moore SJ, Dehui D. Antioxidative and antibacterial activity of the methanol extract of *Artemisia anomala*. *African Journal of Biotechnology*. 2008;7(9):1335-8.
19. Nagalapur SK, Paramjyothi S. In vitro antioxidant activity of launaea pinnatifida cass leaves. *Biogeosciences*. 2010;5(1):105-8.
20. Oladimeji OH, Anwana MA, Attih EE, Effiong DE. 3, 4, 5-trihydroxycyclohexyl methanol-a new reduced derivative from structural activity relationship studies (SARS) on gallic acid. *European Chemical Bulletin*. 2020;9(3):103-6.
21. Jahangir MA, Taleuzzaman M, Alam MJ, Soni A, Beg S. Analytical quality by design for capillary electrophoresis. In *Handbook of Analytical Quality by Design 2021 Jan 1* (pp. 115-132). Academic Press.
22. Taleuzzaman M, Jahangir MA, Chauhan S, Kala C, Beg S. Good laboratory practice and current good manufacturing practice requirements in the development of cancer nanomedicines. In *Nanof ormulation Strategies for Cancer Treatment 2021 Jan 1* (pp. 341-352). Elsevier.
23. Oladimeji OH, Bakare-Odunola MT, Attih EE. Antioxidant Activity of *Acalypha wilkesiana* var. *lace-acalypha* (Muell & Arg.). *African Journal of Pharmacology and Therapeutics*. 2016 Mar 16;5(1).
24. Jahangir MA, Muheem A, Imam SS, Ahmed FJ, Aqil M. *Nigella sativa* Encapsulated Nano-Scaffolds and Their Bioactivity Significance. In *Biomarkers as Targeted Herbal Drug Discovery*. 2021 Jul 4:155-75.
25. Versalovic J, editor. Manual of clinical microbiology. American Society for Microbiology Press; 2011 May 16.
26. Gilani SJ, Jahangir MA, Rizwanullah M, Taleuzzaman M, Shahab MS, Shakeel K, Aqil M, Imam SS. Nano-based therapy for treatment of skin cancer. Recent patents on anti-infective drug discovery. 2018 Aug 1;13(2):151-63.
27. Gilani SJ, Imam SS, Jafar M, Alshehri S, Taleuzzaman M, Jahangir MA. Curcumin Nanomedicine and Their Application in the Management of Disease. In *Biomarkers as Targeted Herbal Drug Discovery*. 2021 Jul 4:43-63.
28. Nia, R., Isolation and characterization of antibacterial constituents from *Calliandra haematocephala* Hassk and *Cissus quadrangularis* L. Ph.D Thesis. Obafemi Awolowo University, Nigeria, 1999, 199p.
29. Adesina SK, Idowu O, Ogundaini AO, Oladimeji H, Olugbade TA, Onawunmi GO, Pais M. Antimicrobial constituents of the leaves of *Acalypha wilkesiana* and *Acalypha hispida*. *Phytotherapy research*. 2000 Aug;14(5):371-4.
30. Oladimeji HO. Chemical and biological studies on *Cyathula prostrata* (L.) Blume. Unpublished. Ph. D. Thesis, University of Uyo, Uyo, Nigeria, 2012b. 2012.
31. Oladimeji OH, Igboasoiki AC. Isolation, characterization and antimicrobial analysis of ethyl gallate and pyrogallol from *Acalypha wilkesiana*

- var. lace-acalypha (Muell & Arg.). African Journal of Pharmacology and Therapeutics. 2014 Sep 30;3(3).
32. Oladimeji OH, Akpan CB. Antioxidant potential of some Nigerian herbal recipes. J. Pharm. Bioresour. 2015;11(2):76-84.
33. Koratoprak, G.S., Antioxidant effects on health. The bright and dark sides. 2022, pp.517-526.
34. Sharma PC, Sharma D, Sharma A, Saini N, Goyal R, Ola M, Chawla R, Thakur VK. Hydrazone comprising compounds as promising anti-infective agents: Chemistry and structure-property relationship. Materials Today Chemistry. 2020 Dec 1;18:100349.
35. Taleuzzamana M, Jahangirb A, Gilania SJ. Quantification and identification of bioactive eugenol in Myristica fragrans seeds using validated high performance thin layer chromatography technique. Pharm. Anal. Acta. 2017;8:E563.
36. Jahangir MA, Muheem A, Anand C, Imam SS. Traditional and modern applications of honey: An insight. Therapeutic Applications of Honey and its Phytochemicals: Vol. 1. 2020:151-69.
37. Muheem A, Shakeel F, Zafar S, Jahangir MA, Warsi MH, Jain GK, Ahmad FJ. Development and validation of stability indicating liquid chromatographic (RP-HPLC) method for estimation of ubidecarenone in bulk drug and formulations using quality by design (QBD) approach. Brazilian Journal of Pharmaceutical Sciences. 2018 Mar 5;53.
-