



Liposomal Nanoformulation of Piperlongumine for Improved Aqueous Solubility and Enhanced Anti-tumour Activity In-vitro

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Abstract

Piperlongumine (PL) is an alkaloid isolated from the Piper species, which can induce apoptosis in cancer cells but has little inhibitory effect on the growth of normal cells. However, its poor aqueous solubility has largely limited its therapeutic applications. To address this issue, we developed liposomal nanoformulations for PL (nanoPLs) in this study. The optimized nanoPLs exhibited a small average diameter in the range of 100-160 nm and narrow size distribution as well as high encapsulation efficiency. NanoPLs also demonstrated excellent storage stability at 4°C for 12 weeks. The apparent solubility of PL in NanoPLs was improved up to 312 folds than in PBS. NanoPLs also markedly increased the cytotoxicity of PL to human prostate cancer PC-3 cells when compared to free PL. NanoPLs represent promising and convenient injectable formulations of PL for further preclinical and clinical investigation.

Keywords: Piperlongumine; Nanoparticles; Liposomes; Cancer; Solubility; Cytotoxicity

Introduction

Piperlongumine (PL, also known as pipartine, CAS#: 20069-09-4, 5,6-dihydro-1-[(2E)-1-oxo-3-(3,4,5-trimethoxyphenyl)-2-propenyl]-2(1H)-pyridinone, Figure 1) is a bioactive alkaloid isolated from black pepper (*Piper nigrum*) and long pepper (*P. longum*) [1]. Previous studies have revealed that PL has different pharmacological effects, including anti-diabetic, anti-fungal, anti-leishmania, anti-inflammatory, anti-platelet aggregation, and anti-cancer activities [2]. Recently, PL has attracted special attention as a potential anti-neoplastic agent because it showed selective cytotoxicity to a plethora of human cancer cells, including pancreatic, breast, lung, colon and bladder cancer, but not to normal cells [3]. Piperlongumine increases the level of reactive oxygen species (ROS) and promotes apoptosis in cancer cells [4]. The selectivity of PL for cancer cells over normal cells is

related to the over-expression of nuclear factor erythroid 2-related factor 2 (Nrf2) in cancer cells [5]. Piperlongumine binds to and inhibits the activity of Nrf2, which is an essential component of the cell to regulate the antioxidants activity, resulting in the reduction of intracellular antioxidants. It was also shown that there are alternative mechanisms of action, including the induction of autophagy in cancer cells by targeting p38 signaling [6] and inhibitory effects on STAT3, which is independent of the ROS mechanism [7].

Despite such appealing potential in cancer treatment, the therapeutic application of PL has yet been embraced mainly due to its strong hydrophobicity and extremely poor aqueous solubility, which has been determined separately by Aodah et al. (0.026 mg/mL) [8] and

Fofaria et al. (0.32 mg/mL) [9]. When dispersed in a physiological solution, PL easily forms aggregates, resulting in the formation of a turbid suspension, but not a solution. Intravenous administration of PL suspension to tumour-bearing animals or to cancer patients will inevitably result in embolism which may be fatal [10, 11].

In addition, the poor aqueous solubility made it impossible for dose escalation in disease treatment. Although Cremophor EL and other vehicles are used in clinic to address the solubility issue of certain therapeutic agents, such as paclitaxel, many problems have been identified with the use of the vehicles, such as peripheral neuropathy, aggregation of erythrocytes, hyperlipidaemia and severe anaphylactoid hypersensitivity reactions [12-16]. Therefore, it is of great importance to develop a delivery system that can significantly increase the solubility of PL to enable parenteral administration in a physiological solution.

Previously, PL has been formulated into chitosan nanoparticles [17], polymeric micelles [5,18], PLGA nanoparticles [19] and nanoemulsions [9] to solve the solubility issue. However, the apparent solubility of PL in these formulations was not reported in any of these nanoformulations. Thus, it is unclear whether these formulations are suitable for intravenous administration. Injectable liposomal nanoformulations for PL delivery have yet been reported. Liposomes are small artificial lipid vesicles consisting of hydrophilic and hydrophobic regions. The aqueous pool of liposomes is surrounded by bilayers of lipids which are composed of natural or synthetic phospholipids. Liposomes have significant advantages in drug delivery to improve the aqueous solubility and stability of therapeutic agents [20-24].

Through the hydrophobic interaction, hydrophobic compounds can be incorporated into the non-polar lipid bilayers of liposomes. Thus, the aqueous solubility of the poorly water-soluble compound can be dramatically improved. In the past several decades, liposomal nanotechnology has helped a number of poorly soluble hydrophobic drugs reach market, such as paclitaxel [25] and irinotecan [26,27]. It also helped numerous promising therapeutic molecules advance to clinical/preclinical investigation, such as ibuprofen [22] and indomethacin [28]. From clinical oncology point of view, by limiting the particle size to approximate 100-200 nm, liposomes can preferentially accumulate in solid tumour tissues after intravenous administration by means of the “enhanced permeation and retention”

(EPR) effect [29-31], and decrease their distribution to normal tissues [32]. In fact, a number of reports have demonstrated enhanced drug concentration in the solid tumour tissue delivered by nanoliposomes than by drug solution [33,34].

Thus, nanoliposomes can further enhance the therapeutic index as well as the antitumor specificity of PL. In this work, we report the preparation, physicochemical characterization, storage stability and cytotoxicity of liposomal nanoformulations of PL (nanoPLs). For the first time, PL was successfully formulated into stable liposomal nanoparticles which markedly improved its aqueous solubility. In addition, nanoPLs exhibited stronger ability to induce cancer cell death compared to free PL.

Materials and Methods

Materials

Phosphatidylcholine (PC), N-[1-(2, 3-Dioleoyloxy) propyl]-N, N, N-trimethylammonium methyl-sulphate (DOTAP) and cholesterol (Chol) were purchased from Avanti Polar Lipids (Alabaster, AL). Piperlongumine (PL) was purchased from Cayman Chemical Co. (Ann Arbor, MI). Phosphate buffer solution (PBS, pH 7.4) was purchased from VWR (Radnor, PA).

Cell culture media (Kaighn's Modification of Ham's F-12k Medium), Trypsin-EDTA Solution, and Penicillin-Streptomycin Solution were purchased from American Type Culture Collection (ATCC) (Manassas, VA). CellTiter 96[®] Aqueous One Solution Cell Proliferation Assay kit was purchased from Promega (Madison, WI).

Cell Culture

Human prostate cancer PC-3 cells were purchased from ATCC (Manassas, VA). The cells were maintained in Kaighn's Modification of Ham's F-12k Medium supplemented with 10% FBS in a humidified CO₂ incubator at 37°C.

Preparation of PL-loaded Liposomal Nanoformulations (NanoPLs)

NanoPLs were prepared using the thin film rehydration method followed by sonication [35]. Briefly, varied amount of PL and lipid materials containing PC, DOTAP and cholesterol at a molar ratio of 60/30/10 were dissolved in 4 mL of chloroform in a round bottom flask. The organic solvent was completely removed on

a rotary evaporator to obtain a thin lipid film in the flask. The dry lipid film was hydrated with PBS to form a crude liposome suspension, and then the suspension was transferred to a glass tube and kept in a refrigerator at 4°C overnight.

The crude liposome suspension was then sonicated at 55°C. The cholesterol percentage remained at 10 % (molar %) while the percentage of DOTAP varied at 30%, 40% and 50% of the total lipid materials. The ratio of PL to total lipid materials was optimized at 1/10, 1.5/10 and 2/10 (molar ratio). For formulations containing 40% and 50% DOTAP, the percentage of PC was lowered to 50% and 40%, respectively. In total, nine different nanoPLs were prepared. Three blank liposomal vehicles which do not contain PL were prepared in the same manner.

Physicochemical Characterization of NanoPLs

The average particle size and size distribution were measured by dynamic light scattering. A Brookhaven NanoBrook 90PLUS particle sizer and zeta potential analyser was used to determine the average size, zeta potential, and polydispersity index (PDI). The samples were diluted in PBS (pH 7.4) by 5 folds prior to the measurement. The intensity of scattered light was detected at 90° angle to an incident beam. All the data analysis was performed in automatic mode.

HPLC Analysis

PL was analysed using high performance liquid chromatography (HPLC). The HPLC system (Agilent Technologies, Santa Clara, CA) consisted of a G4212B diode Array detector, a G1311B/C quaternary pump, two plunger chambers and an Agilent Lab Advisor software. The analysis was performed with a ZOPRAX Extended C-18 (150 mm, 5 microns) column which was maintained at room temperature.

The mobile phase consisted of 70% (v/v) methanol and 30% (v/v) distilled water with the pH being adjusted to 8.1. The flow rate was 1 mL/min and the injection volume was 20 µL. The UV absorbance from PL was measured at 253 nm wavelength [36].

Solubility Study

To determine the solubility of PL in PBS (pH 7.4), excessive amount of PL was added to 1 mL of PBS followed by constant shaking for 24 h at room temperature. Then the PL suspension was visually

checked, and the presence of undissolved PL aggregates was confirmed. PL suspension was centrifuged at 11400 g for 5 min where the undissolved and aggregated PL settled down. The supernatant was diluted in methanol and filtered by syringe filters with the pore size of 0.22 µm prior to analysis by HPLC. For nanoPLs, unencapsulated PL aggregates were removed using the same centrifugation condition. The concentration of PL in the supernatant was determined by HPLC as described above.

Encapsulation Efficiency and Drug Loading Efficiency Determination

NanoPLs were centrifuged at 11400 g for 5 min at 4°C to separate free PL from nanoPLs. The supernatant was carefully taken out from the Eppendorf tube and diluted by methanol, and the concentration of PL in the supernatant was determined by HPLC. The encapsulation efficiency (EE%) was calculated by the following equation:

$$EE\% = \frac{\text{The amount of PL in nanoPLs}}{\text{Total amount of PL added to prepare nanoPLs}} \times 100 \quad (1)$$

The drug loading efficiency (DL%) of nanoPLs was analysed after removing water from the nanoPLs by freeze-drying. The unencapsulated PL aggregates were separated by centrifugation, then nanoPLs supernatant was transferred to a weighted empty glass vial followed by lyophilization. The exact weight of nanoPLs was calculated by deducting the weight of empty glass vial from the weight of the glass vial containing lyophilized nanoPLs. Subsequently, the nanoPLs were dissolved in methanol and the concentration of PL was determined using HPLC. The following equation was used to calculate DL%:

$$DL\% = \frac{\text{The weight of PL in nanoPLs}}{\text{The weight of nanoPLs}} \times 100 \quad (2)$$

Physical Stability Study

NanoPLs were sealed in glass vials and stored at 4°C. The particle size and PDI were analysed every 4 weeks for a period of 12 weeks. EE % and DL % were measured immediately after preparation (week 0) and after 12 weeks (week 12).

In vitro Antitumor Activity

Cytotoxicity analysis was performed using a CellTiter 96® Aqueous Cell Proliferation Assay kit. Briefly,

human prostate cancer PC-3 cells were seeded in a 96-well plate at a density of 1×10^4 cells per well 24 hours prior to the treatment. Then the cells were treated with free PL (in DMSO) or nanoPLs with the final PL concentrations at 3, 6, 12, 18, 24, and 30 μM in each well for 24 hours.

When the treatment was finished, the relative cell viability compared to non-treated cells was measured using the kit according to the manufacturer's protocol. The absorbance was taken at 490 nm using a microplate reader. All experiments were set up in triplicates to determine means and standard deviations. The percentage of viable cells was calculated as follows:

$$\text{Cell viability (\%)} = \frac{\text{Absorbance intensity of treated cells}}{\text{Absorbance intensity of non-treated cells}} \times 100 \quad (3)$$

To determine the half maximal inhibitory concentration (IC₅₀) values of PL and nanoPLs to PC-3 cells, the cells were seeded at 1×10^4 cells per well in a 96 well plate 24 hours prior to the treatment. The cells were exposed to free PL and nanoPLs at different concentrations (0.01, 0.1, 1, 10 and 100 μM) for 24 hours, then the cell viability was determined using the cell proliferation assay kit. The IC₅₀ values of free PL and nanoPLs against PC-3 cells were calculated using GraphPad Prism [37].

Statistical Analysis

Data were presented as mean \pm standard deviation. GraphPad Prism software was used for statistical analysis. For comparison between two groups, linear regression with 95% confidence interval and unpaired two-tailed Students' t test was performed. For comparison of paired data among three or more groups, one-way analysis of variance (ANOVA) followed by a Tukey's post hoc analysis was used. Difference was considered statistically significant when a value of $p < 0.05$.

Results

Characterization of PL-loaded liposomal nanoformulations (nanoPLs)

To prepare nanoPLs, the simple yet classic thin film rehydration followed by sonication method [35] was used. According to this method, PL was first mixed with lipid materials to form a thin and dry film, and then the film was re-dispersed in PBS. Nine different liposomal

nanoformulations containing PL were prepared to investigate the influence of the DOTAP percentage in the lipid materials and the ratio of PL to total lipid materials on the characterization of nanoPLs. The particle size, size distribution and surface charge of nanoPLs are summarized in Table 1. To evaluate the effect of DOTAP% on the physicochemical characteristics of the nanoPLs, the cholesterol % remained unchanged at 10% by molar. Dynamic light scattering data showed that when the ratio of PL to total lipid materials was fixed at 1/10 (molar ratio), increasing the DOTAP percentage resulted in the decrease of average particle size from 160.4 ± 7.7 nm (30% DOTAP, formulation 1C) to 123.1 ± 7.6 nm (40% DOTAP, formulation 2C), and further decrease to 102.9 ± 2.4 nm (50% DOTAP, formulation 3C). The similar decrease was observed when the ratio of PL to total lipid materials was 1.5/10. However, when the ratio of PL to total lipid materials was 2/10, altering DOTAP% did not significantly reduce the particle size of nanoPLs. The surface charge of nanoPLs was also increased from 18.9 ± 9.01 mV (formulation 1C) to 26.9 ± 8.20 mV (formulation 2C), then further to 49.8 ± 15.40 mV (formulation 3C) when the percentage of DOTAP was increased from 30% to 40% and 50%.

When the PC/DOTAP/Chol is fixed at 40/50/10 (molar ratio), lowering the ratio of PL to total lipid materials significantly decreased the average particle size from 263.4 ± 30.2 nm (formulation 3A) to 217.0 ± 16.5 nm (formulation 3B) then to 102.9 ± 2.4 nm (formulation 3C). Formulations 1C, 2C, and 3C were chosen for further characterization as they represented smaller particle size as well as narrower size distribution (Table 1). Using HPLC, the encapsulation efficiency (EE%) of formulation 1C, 2C and 3C was determined to be 93.8 ± 0.04 , 95.2 ± 0.01 , and $97.5 \pm 0.03\%$, respectively, and drug loading efficiency (DL%) of the three formulations to be 2.3 ± 0.03 , 2.6 ± 0.18 , and $1.2 \pm 0.02\%$, respectively (Table 2).

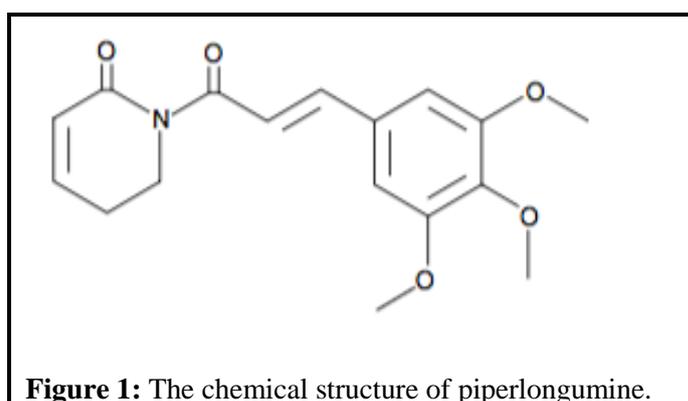


Figure 1: The chemical structure of piperlongumine.

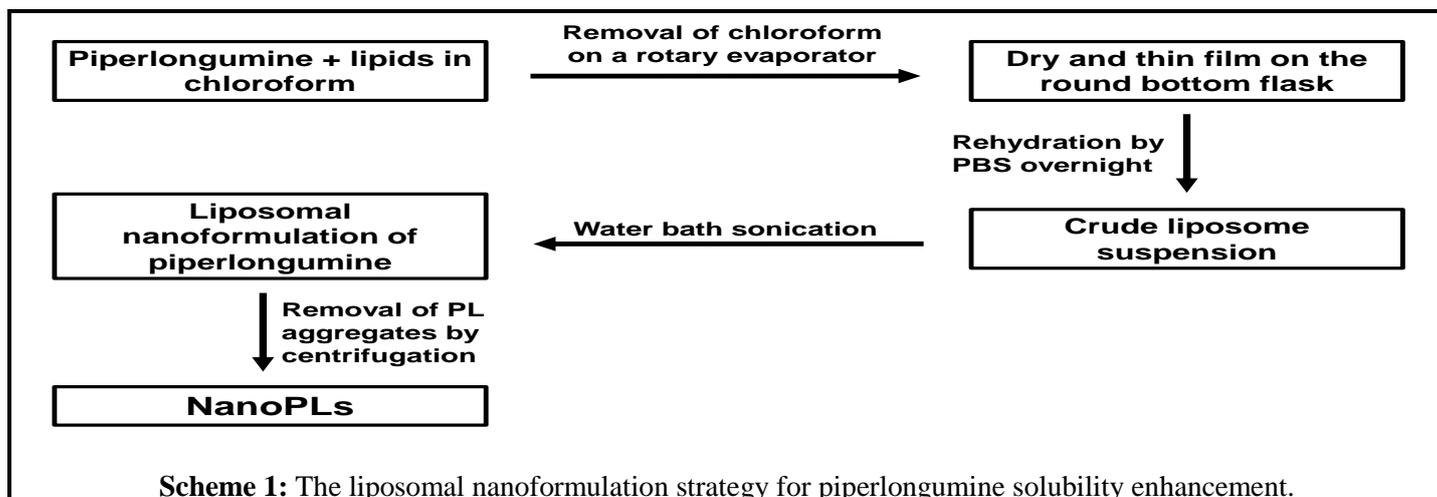


Table 1: NanoPLs: formulation composition, particle size and surface charge.

Formulation ID	PC/DOTAP/Chol (Molar ratio)	PL to total lipid ratio (Molar ratio)	Size (d. nm)	Polydispersity index (PDI)	Zeta potential (mV)
1A	60/30/10	02-10	268.1 ± 52.8	0.34 ± 0.03	9.3 ± 3.30
1B	60/30/10	1.5/10	280.8 ± 7.6	0.36 ± 0.01	27.7 ± 1.40
1C	60/30/10	01-10	160.4 ± 7.7	0.21 ± 0.01	18.91 ± 9.01
2A	50/40/10	02-10	277.9 ± 32.7	0.36 ± 0.02	24 ± 0.60
2B	50/40/10	1.5/10	247.6 ± 20.9	0.33 ± 0.20	32.3 ± 4.40
2C	50/40/10	01-10	123.1 ± 7.6	0.24 ± 0.02	26.9 ± 8.20
3A	40/50/10	02-10	263.4 ± 30.2	0.32 ± 0.01	37.8 ± 4.90
3B	40/50/10	1.5/10	217.0 ± 16.5	0.34 ± 0.01	29.6 ± 16.40
3C	40/50/10	01-10	102.9 ± 2.4	0.29 ± 0.02	49.8 ± 15.40

Table 2: The encapsulation efficiency (EE%) and drug loading efficiency (DL%) of NanoPLs.

Formulation ID	Time 0		12 weeks	
	EE%	DL%	EE%	DL%
1C	93.83 ± 0.04	2.30 ± 0.03	94.09 ± 0.02	2.32 ± 0.05
2C	95.23 ± 0.01	2.38 ± 0.18	93.61 ± 0.01	2.33 ± 0.02
3C	97.46 ± 0.03	1.19 ± 0.32	95.45 ± 0.03	1.03 ± 0.14

Solubility of PL in liposomal nanoformulations

The apparent solubility of PL in PBS and in NanoPL 1C, 2C, 3C was determined after separating PL aggregates from nanoPLs by centrifugation. There were many undissolved flakes when the excessive amount of PL was dispersed in PBS. After centrifugation, a white pellet at the bottom of the Eppendorf tube was easily seen (Figure 2A, left). HPLC analysis data showed that the solubility of PL in PBS at pH 7.4 is only 0.02 ± 0.01 mg/mL. On the contrary, nanoPLs were almost clear and well-dispersed formulations. No visible pellets were observed after centrifugation (Figure 2A, right). The solubility of PL in 1C, 2C, and 3C formulations was 5.14 ± 0.38 mg/mL, 6.25 ± 0.44 mg/mL and 4.64 ± 0.25 mg/mL, respectively (p<0.05, Figure 2B). The apparent solubility of PL in NanoPL 1C, 2C and 3C exceeded the

intrinsic solubility of free PL in PBS by 257, 312 and 232 times, respectively.

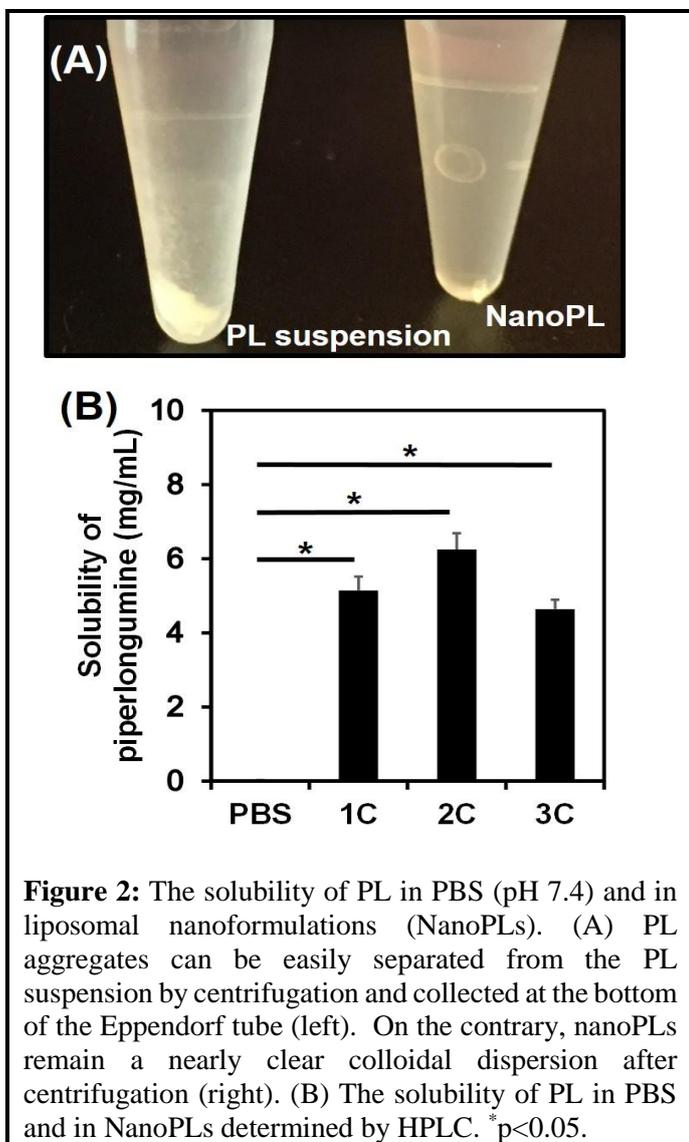


Figure 2: The solubility of PL in PBS (pH 7.4) and in liposomal nanoformulations (NanoPLs). (A) PL aggregates can be easily separated from the PL suspension by centrifugation and collected at the bottom of the Eppendorf tube (left). On the contrary, nanoPLs remain a nearly clear colloidal dispersion after centrifugation (right). (B) The solubility of PL in PBS and in NanoPLs determined by HPLC. *p<0.05.

Storage stability of nanoPLs

The storage stability of NanoPL 1C, 2C, and 3C were evaluated at 4°C for 12 weeks. The average particle size and size distribution were measured at 4, 8 and 12 weeks. The EE% and DL% were determined after 12 weeks. NanoPL 1C, 2C, and 3C all demonstrated high stability during storage. No significant change in particle size, size distribution, EE% or DL% was observed during the test (p>0.05, Figure 3A, 3B and Table 2).

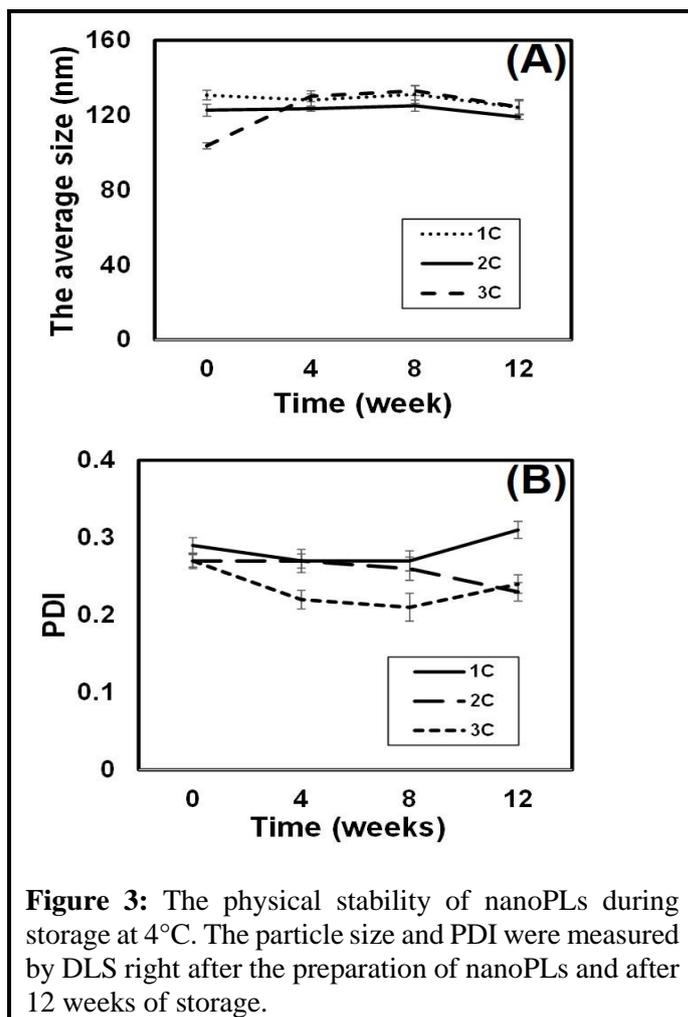


Figure 3: The physical stability of nanoPLs during storage at 4°C. The particle size and PDI were measured by DLS right after the preparation of nanoPLs and after 12 weeks of storage.

In vitro antitumor activity against human prostate cancer PC-3 cells

The *In vitro* antitumor effects of nanoPLs were evaluated using human prostate cancer PC-3 cell line. In order to examine the possible toxicity from the lipid materials, three empty liposomal vehicles containing the same lipid composition as of formulation 1C, 2C and 3C were prepared. PC-3 cells were exposed to nanoPLs at the final PL concentration of 24 µM. Control groups were treated by empty liposomal vehicles at the same lipid concentrations. As shown in Figure 4, after 24 hours of treatment, nanoPLs exhibited strong cell killing ability by reducing the cell viability to 53% or less, whereas empty liposomal vehicles of 1C and 2C only slightly affected cells by reducing the cell viability to 88.72 ± 7.4% and 87.4 ± 5.4%. However, after the treatment by empty liposomal vehicles of 3C, the cell viability decreased to 67.54 ± 1.95%, which indicated that the liposomal vehicle used in NanoPL 3C indeed was toxic.

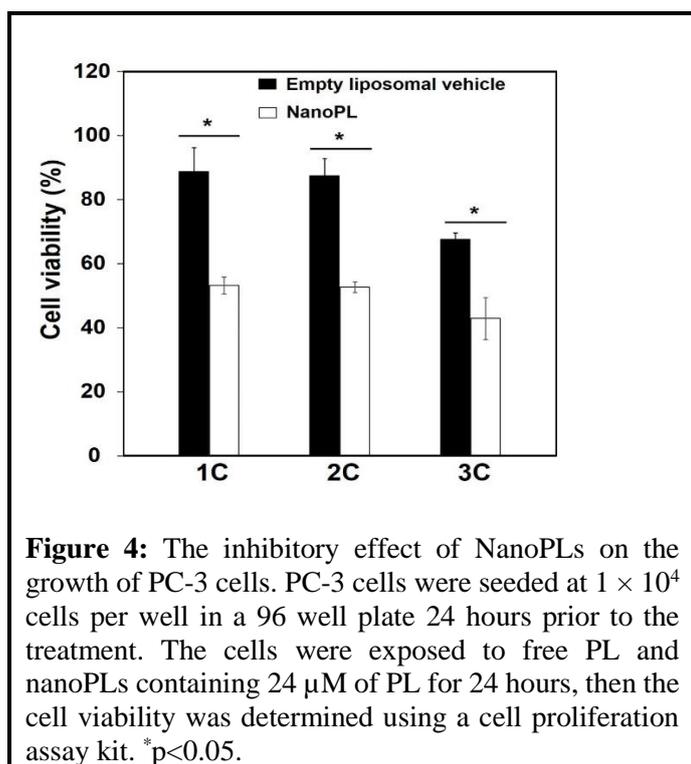


Figure 4: The inhibitory effect of NanoPLs on the growth of PC-3 cells. PC-3 cells were seeded at 1×10^4 cells per well in a 96 well plate 24 hours prior to the treatment. The cells were exposed to free PL and nanoPLs containing 24 μM of PL for 24 hours, then the cell viability was determined using a cell proliferation assay kit. * $p < 0.05$.

After confirming the anti-tumour activity of nanoPLs, we further investigated the dose response. PC-3 cells were treated by free PL and nanoPLs at escalated concentrations from 3 to 30 μM . We found that the relative percentage of viable cells treated by formulations 1C, 2C, and 3C was significantly lower than that of cells treated by free PL ($p < 0.05$, Figure 5).

At the lower dose of 3 μM , the viability of the cells treated by free PL was $91.2 \pm 0.93\%$. At the same dose, the cell viability with liposomal formulations 1C, 2C, and 3C were $93.34 \pm 4.8\%$, $87.98 \pm 6.19\%$, and $87.76 \pm 3.92\%$, respectively.

The cell viability gradually decreased with the increase of PL concentration. Clear dose-dependent response was observed for both free PL and nanoPLs (Figure 5). After the treatment by 30 μM of free PL, the viable cells were $86.93 \pm 3.99\%$. But the percentage of viable cells was significantly decreased the treatment by nanoPLs at the same dose level of PL. Specifically, the viability were $47.05 \pm 4.83\%$, $41.56 \pm 3.94\%$, and $28.92 \pm 1.22\%$, after the treatment by 1C, 2C, and 3C formulations, respectively. To best describe the anti-tumour activity, the IC₅₀ values were calculated for nanoPLs 1C, 2C, 3C and free PL. As shown in Table 3, the IC₅₀ of formulations 1C, 2C, 3C were $35.15 \pm 3.27 \mu\text{M}$, $29.49 \pm 0.03 \mu\text{M}$, and $23.74 \pm 2.05 \mu\text{M}$, respectively, which

were significantly lower than that of free PL, which was $45.68 \pm 5.02 \mu\text{M}$.

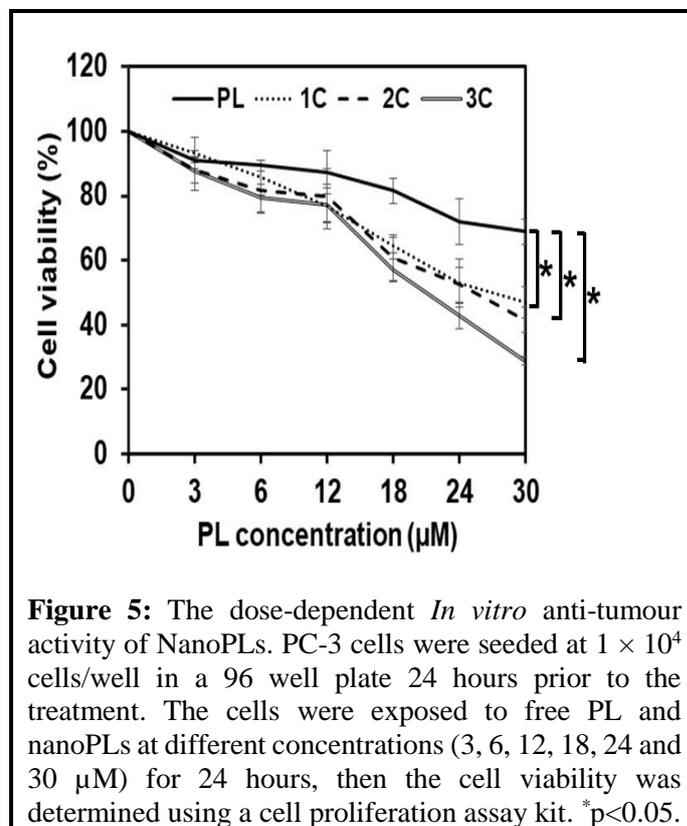


Figure 5: The dose-dependent *In vitro* anti-tumour activity of NanoPLs. PC-3 cells were seeded at 1×10^4 cells/well in a 96 well plate 24 hours prior to the treatment. The cells were exposed to free PL and nanoPLs at different concentrations (3, 6, 12, 18, 24 and 30 μM) for 24 hours, then the cell viability was determined using a cell proliferation assay kit. * $p < 0.05$.

Table 3: The IC₅₀ of free PL and NanoPLs to PC-3 cells. [Note: * $p < 0.05$ versus free PL. ** $p < 0.01$ versus free PL. *** $p < 0.005$ versus free PL].

Sample	IC ₅₀ (mM)
Free PL	45.68 ± 5.02
1C	$35.15 \pm 3.27^*$
2C	$29.49 \pm 0.03^{**}$
3C	$23.74 \pm 2.05^{***}$

Discussion

With the solubility in PBS (pH=7.4) at only $0.02 \pm 0.01 \text{ mg/mL}$ (Figure 2B), PL is a “practically insoluble (PI)” compound based on the solubility classification in the US Pharmacopeia. Although PL has demonstrated several pharmacological activities, especially in cancer treatment, this poor aqueous solubility has largely impeded its clinical investigation and application. Previously reported polymeric nanoparticles and micelles [5,17-19] are valuable tools to improve PL’s solubility, but none of the polymer materials have been approved by the US FDA for intravenous administration. The safety and biocompatibility of these

polymers in human is still unclear. Liposomal nanoparticles are apparently advantageous over polymeric nanoparticles and polymeric micelles due to the excellent biocompatibility and safety record of lipid materials in human [24]. For example, liposomal formulation Doxil[®] (liposomal doxorubicin) and Marqibo[®] (liposomal vincristine) were approved by the US FDA because both products are able to enhance the apparent solubility and therapeutic efficacy of the drug in cancer. Therefore, the primary objective of this study was to develop a liposomal nanoformulation, in which PL is sufficiently soluble and suitable for intravenous administration. We report for the first time that liposomal nanoformulations dramatically improved the aqueous solubility of PL with a very high encapsulation efficiency (Table 2). Specifically, formulation 2C raised the solubility of PL to 6.25 ± 0.44 mg/mL. Because the volume of general intravenous injection to each mouse is limited to 100 μ L or less, this markedly elevated PL aqueous solubility makes it very convenient for *In vitro* anti-tumour experiments using tumour-bearing mouse models, which we have already planned for. It is important to note that even the commercial product Doxil[®] can only raise the doxorubicin concentration to 2 mg/mL [32] and Marqibo[®] can only solubilize vincristine to 0.16 mg/mL [38]. Therefore, the aqueous solubility problem of PL is successfully addressed by using liposomal nanotechnology.

The particle size, size distribution and storage stability are crucial factors for the successful application of liposomal nanoparticles. During the formulation development, the influence of the percentage of DOTAP lipid on the particle size and stability of nanoPLs was evaluated. Our results have shown that higher percentage of DOTAP in the formulation helped reduce the size and enhance the stability of liposome formulations, probably by increasing the fluidity of lipid membrane. This is consistent with the results reported by Campbell and co-workers [39,40]. In addition, DOTAP contributes positive surface charge to nanoPLs, resulting in elevated electro-repulsive tension and reduced the tendency of collision between liposomal nanoparticles. Another key parameter influencing the particle size is the ratio of PL to total lipids. Due to the strong hydrophobicity, PL molecules have to reside in the lipid bilayer of liposomes in order to avoid forming aggregates. When the ratio of PL to total lipids is high, e.g. at 2/10, there are more PL molecules in the lipid bilayer, which increased the rigidity and reduced the curvature of the lipid bilayer. Therefore, the particles prepared at 2/10 ratio are relatively larger than at 1/10 ratio. The excellent stability of nanoPLs should also be

attributed, at least in part, to the addition of natural lipid cholesterol [41]. Overall, by rationally controlling the lipid components, particle size and surface charge, nanoPLs (1C, 2C and 3C) developed in the study demonstrated excellent storage stability (Table 2 and Figure 3).

In the last part of this study, the *In vitro* anti-tumour activity was investigated and the IC₅₀ of free PL and nanoPLs to PC-3 cells were determined. Because nanoparticles greater than 200 nm in diameter are usually prone to aggregate during storage in buffer, are rapidly cleared in the blood stream, and are less efficient in passive accumulation in solid tumour tissues [42,43], we selected formulations with the average particle size below 200 nm for cytotoxicity studies. Compare to free PL, nanoPLs exhibited a significantly increased cytotoxicity ($p < 0.05$) (Figures 4 and 5). The strongest antitumor activity was achieved with formulation 3C. The enhanced cytotoxicity can be accounted for the positive surface charge of nanoPLs conferred by DOTAP. Since mammalian cell membrane has a negative surface charge, positively charged liposomes have a high affinity to the cells based on electrostatic interaction [39,44,45].

We investigated whether the enhanced cell killing activity of nanoPLs was caused by the cytotoxicity of liposomal vehicles. Three different empty liposomal formulations were prepared using the same lipid components and composition used in nanoPLs 1C, 2C and 3C, and then PC-3 cells were treated with the empty liposome controls at the same lipid concentration. As shown in Figure 4, empty liposomal vehicles used in formulation 1C and 2C, which contain relatively lower percentage of DOTAP, showed little toxicity to PC-3 cells.

The results indicated that formulations 1C and 2C induced cancer cell death indeed by PL, very likely through enhanced intracellular delivery by cationic liposomes. It should be pointed out that when the DOTAP content was increased to 50%, empty liposomal vehicles used in nanoformulation 3C did show obvious toxicity to PC-3 cells. We further demonstrated that nanoPLs could elicit a strong anti-tumour activity in a dose-dependent manner to prostate cancer PC-3 cells (Figure 5). In addition, the IC₅₀ values of nanoPLs were significantly lower than that of free PL (Table 3). Therefore, nanoPLs 1C and 2C appear to be better choice for *In vitro* investigation in the future.

Conclusion

In this study, the first liposomal nanoformulation for the natural active compound PL was developed. The aqueous solubility issue of PL was successfully solved using liposomal nanotechnology. NanoPLs demonstrated excellent storage stability and strong anti-tumour activity *In vitro*. The results of this study warrant further investigation of nanoPLs in animal models of human prostate cancer as well as other cancer types. NanoPLs provide a “soluble” form of PL in aqueous solution, which can be conveniently prepared by most of the research laboratories and easily used by researchers from a variety of therapeutic areas, such as oncology, cardiology and inflammation, etc. Our results showed that nanoPLs have the potential to dramatically boost the preclinical and clinical investigation of PL.

Conflict of Interest

The authors declare that there are no conflicts of interest.

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